Ensure Accurate and Successful BOD Measurements with Electrochemical or Optical Sensors

Biochemical Oxygen Demand (BOD) Testing
Technical Note T605-02

YSI created this document to address some common problems that are encountered when conducting Biochemical Oxygen Demand (BOD) testing. There can be confusion about BOD and its relationship with dissolved oxygen (DO) readings. Even though the end result of BOD testing is a final BOD reading, the calculations are all based on DO readings. Therefore, DO readings must be as accurate as possible to ultimately get accurate BOD results.

YSI has been a leader in the field of DO measurements for 55+ years. Several of YSI’s BOD probes incorporate the Clark Polarographic, membrane covered, sensor technology (probes include the 5905, 5010, 200-BOD, ProBOD and 5750) while the YSI ProOBOD™ used with the ProODO and MultiLab instruments utilizes membrane-less optical DO technology.

The sensors do not measure the concentration of oxygen but, instead, the sensor measures the partial pressure of dissolved oxygen in % saturation. We know this because water can hold more oxygen in fresh water than in sea water, however, the sensors raw output signal is identical in both media since the pressure is identical in both media.

The instrument then calculates the concentration from the % saturation value (pressure value), the sample temperature, and a measured or entered salinity correction value. The calculation is based on the Oxygen Solubility chart found in Standard Methods. The instrument software automatically calculates the mg/L values.

For more in depth information about dissolved oxygen, please visit YSI.com\weknowDO. For more information on BOD, please visit the YSI BOD parameter page.

There are several factors that can affect DO readings, including the probe care and use, calibration techniques, and other laboratory practices. At some point many labs, if not all, have some issue with their BOD results from blanks coming out too high or poor glucose-glutamic acid (GAA) results. The steps in correcting these issues can often be numerous, time-consuming and outright frustrating. One easy step to take out of the equation is the functionality of the instrument and probe. 

**Probe Use and Care - Electrochemical**

In order for your DO reading, and ultimately your BOD result, to be as accurate as possible the following issues surrounding the use and care of the probe must be addressed.

1. The electrolyte solution should be fresh. Once it has been hydrated, the shelf life of the solution is one year. It is best to allow the electrolyte solution a full 24 hours after mixing before first use to ensure proper mixing of the solution. Care should also be taken to ensure that the correct solution is being used for the correct membrane. The solution you use for your field DO probe may not be the same solution needed for your lab BOD probe and vice versa. Follow the instructions on the electrolyte bottle or download the membrane selection guide at YSI.com\weknowDO (located under the Guides tab).

2. Membranes must be installed correctly. Bubbles, wrinkles, and obvious tears and/or holes can create problems. However, occasional cosmetic blemishes of the membrane are normal and generally do not affect the successful operation and functionality of the probe. The
cap membranes rarely have any issues when stored and used properly.

Membrane caps are a much easier option than the stretch over style membranes common on older DO field probes. The cap membranes have been available for years and are capable of being changed within seconds. Some manufacturers may try to state that membrane changes are costly and cumbersome but it’s as simple as unscrewing the old cap and screwing on a new one. Proper membrane cap installation is shown below in Figure 2.

Instructions for Texturing the Cathode - Electrochemical

The cathode (gold circle) on most YSI probes can be restored to its normal polish by wet sanding the electrode. To sand the cathode, place a 400 grit wet/dry sanding disc, face-up in the palm of your hand. Next, touch the cathode to the sanding disc and twist the probe 3-4 times on the disc. After sanding, rinse the electrode with clean water and inspect to see that the original matte finish is restored. (The gold cathode should have many fine scratches on the surface.) If the cathode remains tarnished it may need to be returned to YSI for service.

3. The gold cathode should be bright and have fine scratches covering the entire surface (See Figure 3). Do not attempt to create a “glass smooth” surface on the cathode, because the absence of scratches will prevent accurate readings from being measured. The gold cathode can also become plated with silver from the anode and tarnished from use with wrinkled membranes or after having contact with known interfering gases such as hydrogen sulfide.

In order to service the gold cathode, follow the instructions below using the sanding disk provided in the Cap Membrane Kit that was included with your YSI BOD probe.

Instructions for Cleaning the Anode - Electrochemical

The silver anode, from which the current is sent out, should be light in color or silver-looking (see Figure 3). A dull, tarnished anode can result in inaccurate readings due to the resultant lower probe current. Some dissolved oxygen instruments may indicate this problem by displaying an error message during calibration.

In the past, a typical practice was to leave the dissolved oxygen instrument turned on overnight. However, the entire time the instrument is turned on it is sending voltage through the anode and the life of the probe is being unnecessarily reduced. The dissolved oxygen instrument should be
turned off after use in the evening and powered up in the morning, allowing the instrument enough time to warm up and stabilize before calibration.

The anode may periodically need “cleaning” or servicing in order to remove the tarnish that appears. The tarnish results from wrinkled or loose membranes, interfering gases, and heavy probe usage. The time period required between anode cleanings depends on the different uses and applications. However, one should only clean the anode when it is absolutely necessary, and avoid cleaning too frequently. The probe can operate perfectly well even with a tarnished look. When you begin to have difficulty calibrating, or experience drifting, jumpy readings, first change the membrane to eliminate the membrane and solution as the cause. Then, if still necessary, clean the anode following the instructions below.

1. To physically clean the electrodes on the DO membrane cap-style probes, you can wet sand the anode with 400 grit sand paper. Simply wrap the sanding disc around the anode (silver portion making up the shaft of the sensor) and, in a helical fashion, twist the sanding disc down and around.

2. To chemically clean the electrodes, soak the probe (without the membrane) in a 14% Ammonium Hydroxide solution for 2-3 minutes. A solution of 3% can also be used (household cleaners with ammonia are about 3%) to soak the probe overnight. After soaking, rinse the probe EXTREMELY WELL under running tap water for 4-5 minutes. (You can smell the probe tip to make sure there is no residual ammonia left). Then rinse with deionized water, rinse with KCl electrolyte solution, and install a new membrane using fresh KCl solution. You may also want to physically clean the anode with the sanding disc after the ammonium soak.

Dissolved Oxygen and Temperature - Electrochemical

DO values are based on temperature compensation, so it is crucial for the temperature readings of your probe to be accurate. Although it rarely occurs, if a thermistor goes out of tolerance, your temperature readings will be obviously incorrect. Temperature affects DO readings in several ways. First, it changes the permeability of the membrane itself. As the temperature increases, the permeability of the membrane also increases. As the temperature decreases, the permeability of the oxygen through the membrane decreases. Secondly, temperature affects the amount of oxygen that can be dissolved in water. As the temperature increases, the oxygen saturation of the water decreases. Since YSI's dissolved oxygen instruments have automatic temperature compensation, it is important to occasionally check the instrument's temperature accuracy.

Calibration - Electrochemical

The next most common problem happens during calibration. An incorrectly calibrated instrument will result in incorrect DO readings. Improper calibration appears to be the main cause for many erroneous DO readings. The effects of an improperly calibrated instrument can result in odd values - high or low. It can also result in DO values of your blank water being “higher” on day 5 than on day 1. YSI encourages an air-saturated calibration procedure for DO probes and instruments. Compensating for altitude or barometric pressure is also important and all of YSI’s instruments have the capability of compensating. Your data is only as good as your calibration.

1. Read the Operations Manual that is provided with both the YSI Dissolved Oxygen Instrument and the probes.

2. Allow proper warm up time for the instrument. This is especially important for new membranes and probes. New membranes need a short “break-in” period of approximately 30 minutes before taking readings. A new probe should be “broken-in” overnight.

3. Allow the temperature in the bottle time to equilibrate with the room temperature if there is a large temperature gradient.

4. Compensate for altitude or barometric pressure. Note that calibrating using only altitude does not account for local weather conditions. Weather variations are typically +/- 1% from week to week. Extreme weather can cause variations up to four times larger. If it was clear and sunny under a high pressure system when the initial DOs were taken, then altitude calibration can cause all readings that day to be off about 1%. Conversely, if it is rainy under a low pressure
system when the final DOs are read, then all readings that day may be off about 1% in the other direction. The additive effect in this scenario is 2%, and 2% of 9 mg/L is 0.18 mg/L! This value approaches or exceeds the error budget for this method and does not account for any degree of accuracy on the system! Remember barometer readings received from the local airport are usually corrected for sea level. Inches of mercury multiplied by 25.4 will result in mm of mercury. You must then determine the true barometric pressure. An approximate formula for this “uncorrection” is below:

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\text{True BP} = [\text{corrected BP mmHg}] - [2.5 \times (\text{local altitude in ft. above sea level})/100]
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Several instruments have internal barometers so you may have to occasionally verify that the internal barometer is reading accurately and calibrate it only if needed.

5. Calibrate in a 100% saturated air environment. Make sure there are no water droplets on the membrane. Water on the membrane during air calibration will cause a low calibration reading due to the oxygen in the droplets being consumed. Excess water may be removed by shaking the probe or dabbing them with a laboratory-grade cleaning tissue.

In addition, make sure the calibration environment is clean. Change the water in the BOD bottles occasionally and use a clean bottle. Using water in the calibration bottle that has “stuff” growing in it will not yield consistent calibrations. See Figure 6.

6. One successful calibration per day is all that is needed. Avoid multiple or back to back calibrations.

Theory - Optical

Briefly, this technology utilizes an optical, luminescent DO sensor in order to determine dissolved oxygen values. The optical field probes and OBOD™ laboratory probe both work in the same manner.

The probe measures dissolved oxygen by emitting a blue light of the proper wavelength that causes the dye in the sensing element to luminesce, or glow red. Oxygen dissolved in the sample continually passes through the diffusion layer to the dye layer, affecting the luminescence of the dye both in intensity and lifetime. The YSI sensor measures lifetime of the dye’s luminescence as it is affected by the presence of oxygen with a photodiode (light detector) in the probe and compares that reading to a reference. See Figure 7.

To increase the accuracy and stability of the measurement, the sensor also emits a red light that is reflected by the dye layer back to the photodiode in the sensor. The sensor measures the reflected light and uses that reading as the reference value for comparison to the previously measured lifetime luminescent value. The lifetime of the luminescence from excitation by the blue light is compared to that of the reference value (red light) and a stable dissolved oxygen concentration is calculated by the probe.

Although the accuracy of an optical sensor’s measurement is not dependent on flow, the YSI ProOBOD™ optical BOD probe does have a stirrer. This stirring ensures a truly representative sample in the BOD bottle and YSI scientists have determined that flow improves the optical sensors response time.

There are several factors that can affect DO readings, including the probe care and use, calibration techniques, and other laboratory practices. At some point many labs, if not all, have some issue with their BOD results from
blanks coming out too high to poor GAA results. The steps in correcting these issues can often be numerous, time-consuming and outright frustrating. One easy step to take out of the equation is the functionality of the instrument and probe.

**Probe Use and Care - Optical**

In order for your DO reading, and ultimately your BOD result, to be as accurate as possible the following issues surrounding the use and care of the probe must be addressed.

1. Clean the probe and sensing element. In order to clean the optical DO sensors, rinse it with clean water and wipe with a lint free cloth. If necessary, use a mild detergent. Do not use alcohols or other organic solvents that may deteriorate the sensing element.

2. The sensing element should be replaced about once per year but may last longer. It should also be replaced if it is cracked or damaged. While changing the sensing element, rinse the optics with clean water and then wipe the optics with a lint free cloth or lens tissue. The instruction sheet that is shipped with the replacement sensing element includes calibration coefficients specific to the sensing element. For highest accuracy, these coefficients should be entered by the user into the instrument following the instructions provided.

**Dissolved Oxygen and Temperature - Optical**

DO values are based on temperature compensation, so it is crucial for the temperature readings of your probe to be accurate. Although it rarely occurs, if a thermistor goes out of tolerance, your temperature readings will be obviously incorrect and you may need to replace the sensing portion of your OBOD probe.

Temperature affects the amount of oxygen that can be dissolved in water. As the temperature increases, the oxygen saturation of the water decreases. Since YSI’s dissolved oxygen instruments have automatic temperature compensation, it is important to occasionally check the instrument’s temperature accuracy.

This temperature dependence is removed by proprietary algorithms in the system software. As for any oxygen probe, the mg/L concentration is calculated from the sensor’s % saturation reading, temperature, and salinity after calibration of the system using barometric pressure.

**Calibration - Optical**

The next most common problem happens during calibration. An incorrectly calibrated instrument will result in incorrect DO readings. Improper calibration appears to be the main cause for many erroneous DO readings. The effects of an improperly calibrated instrument can result in erratic readings including unstable, jumpy, or odd values. It can also result in DO values of your blank water being “higher” on day 5 than on day 1. YSI encourages an air-saturated calibration procedure for DO probes and instruments. Compensating for altitude or barometric pressure is also important and all of YSI’s instruments have the capability of compensating.

1. **Read the Operations Manual** that is provided with both the YSI Dissolved Oxygen Instrument and the probes.

2. **Inspect.** Visually inspect the sensor cap for biofouling or large scratches. Biofouling could consume or produce oxygen thereby affecting the calibration. Large scratches in the paint surface could affect the calibration as well.

3. **Ensure there aren’t any water droplets** on the sensing element or thermistor and dab them off or shake them off if there are. Also ensure that the temperature is stable - since you are in the laboratory this should already be good.

4. **Compensate for altitude or barometric pressure.** The ProODO and all MultiLab instruments have an internal barometer. They will automatically use this value as part of the calibration routine.

5. **Compensate for salinity.** This is the value of the water you will be testing. Make sure the salinity value is set to the value of the samples in the BOD bottles. If this value is 0 then you can set it the first time and forget it.

6. **Calibrate in a 100% saturated air environment.** The calibration should take place in the air in the BOD bottle and not submerged in the water in the bottle.

Figure 8. The YSI MultiLab 4010-3 can measure DO with an OBOD sensor, pH, temperature, ammonia, and many other parameters. The instrument also features a built-in barometer.
Other Factors Affecting DO Readings

There are other factors that may affect your BOD readings or your blanks. Please consider the following:

- **Cleanliness.** Cleanliness can be very difficult to track. This is usually what most labs begin to examine when they have blank depletions coming out too high. Many items could be dirty including the water itself or anything that comes in contact with the water from the time it comes out of the still until its final DO is measured.

- **Supersaturation.** DO may out-gas during the five days giving artificially high depletions and BODs. This is most common with cold river water samples collected during the winter. They may not have been supersaturated in the field, but become supersaturated when they warm up in the lab. A BOD test cannot begin with a supersaturated sample. This limits the total DO available for the test to 9 mg/L.

- **Dilution Water.** The dilution water used is also very important to accurate data. Make sure the dilution water carboy is cleaned properly. Another source of water could be used if there are issues to help determine differences. In addition, DI and RO water systems may need to have the resin cartridges and/or membranes replaced. Discard unused dilution water and thoroughly triple-rinse the carboy with DI water.

- **Bubbles.** Oxygen does not dissolve well in water and a small bubble of air has a large supply of oxygen compared to the amount that can be dissolved in the water sample. An air bubble that is only 0.1 mL in volume contains about 0.6 mg of oxygen at standard temperature and pressure. In a 300 mL bottle over 5 days, this small air bubble can raise the final DO by 1.8 mg/L.

- **Toxic Material.** Enough toxic material can ensure that there is no depletion at all over the five days. More commonly, there is only enough toxic material to weaken the bacteria, causing an artificially low depletion. Sometimes you can detect the presence of toxins by examining how the BOD varies from one dilution to the next in the same sample. If more dilute bottles have greater BODs, that may indicate that those bottles have less toxic material and biological activity is not suppressed as much.

- **Seeding.** Seeding can also lead to difficulties. Seed compensation must be between 0.6-1.0 mg/L. These limits can be difficult to meet if your influent varies from day to day. Some analysts take their seed material from their influent, but allow it to age for a day. Others use material from some intermediate point in the process, for example, the clarifier. Many people have also had good results with commercially available seed material. Seeding a sample solves one problem but causes another. Since we are trying to measure the BOD of the sample, not the BOD of the sample and the seed material together, another sample of the seed material alone must be run to separate the information. This is the Seed Control Sample.

- **Non-linearity.** A bottle that is twice as dilute may or may not actually deplete half as much DO. Seed calculations assume linearity. Seeded samples are therefore slightly more vulnerable to problems. Ideally, a bottle will deplete roughly half of the DO available, about 4-5 mg/L during the 5 day test. To set the dilutions for this ideal depletion, analysts must know the answer before they start. Fortunately, samples usually do not change very much from day to day and analysts confidently choose dilutions that will keep them within method. Still, there is variation and to compensate for it analysts usually set up 2-3 different dilutions to ensure that at least one of them is within method. Occasionally, there will be samples that have not been done before. If analysts have no idea what the BOD is, they will set up many dilutions to get a large spread. This commonly occurs with industrial samples.

Taking into account all of the possible factors that could lead to erroneous readings, it is imperative for laboratories to become thoroughly knowledgeable with the dissolved oxygen instrumentation they are using. An important QA/QC procedure for individual laboratories to consider is if there are multiple users operating the instruments. Reading the Operations Manuals and following the stated practices along with establishing Standard Operating Procedures (SOPs) will help ensure proper usage.

Once these procedures are established, the repeatability of successful readings will greatly increase. If only one of the above stated problems occurs, you may not experience problems. However, if several of the problems occur at the same time, the likelihood of erroneous readings is much greater.

There may still be occasions when final blanks come out higher than the initials but the likelihood of this occurrence should decrease by following and understanding this document.

For additional information, please contact YSI:
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Resources:
Brake, Perry F. and Raynovic, Michael A. “Laboratory Testing for BOD and CBOD.” NCL of Wisconsin, Inc.